

NEW

C Y T O Q U B E TM

Light-Sheet Microplate Cytometer
C15200-01RGBU



Next Generation 3D Screening

HAMAMATSU
PHOTON IS OUR BUSINESS

Advancing cell-based assays and drug screening with CYTOQUBE[®]

CYTOQUBE[®] is a Light-Sheet Microplate Cytometer that performs high-speed fluorescence imaging and analysis for both 2D and 3D cell-cultured samples. It is capable of scanning 1536, 384 and 96 well microplates, acquiring whole-well 3D fluorescence images.

During the image acquisition process, fluorescence intensity and 3D morphological information of the objects (cells, spheroids, or organoids) are analyzed, and the results are displayed upon completion of the scan.

Our unique patented technology, Zyncscan[®], enables high-throughput cell assays with ease of use and higher reliability.

It contributes to the efficiency of drug screening, toxicity/safety evaluation, and translational research using cells, spheroids, organoids, and bioprinting samples.



Fast 3D fluorescence imaging

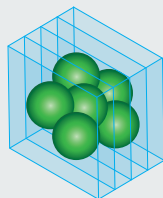


Image acquisition and analysis for 2D and 3D samples (Observable height range: 400 µm or less)

High-throughput analysis



Get 3 color results in

21 min/plate

- The same time applies for all plate formats: 1536, 384, 96.
- Analysis based on 3D imaging is performed in parallel
- Whole-well imaging (up to 400 µm in height from the bottom of the well)

Key challenges addressed with CYTOQUBE[®]

Focus adjustment

- Need to reduce focus adjustment time
- Need to ensure all samples are in focus
- Need to reduce photobleaching effect

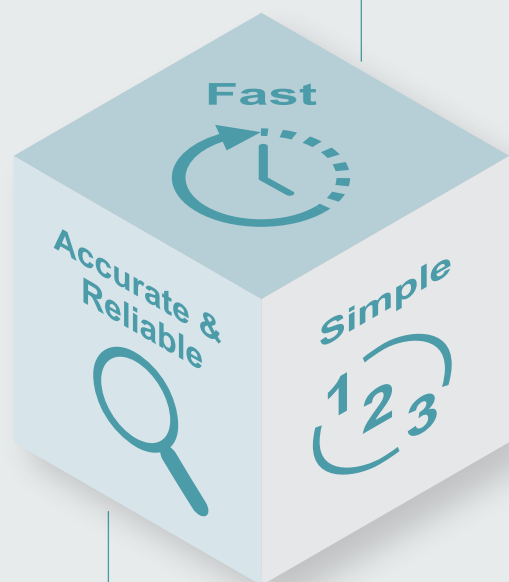
2D / 3D Imaging

- Need to perform 2D/3D imaging and analysis in high throughput
- Need to image cells in U-bottom plates
- Need to reduce photobleaching effect in 3D imaging

Analysis

- Need analyzed results faster and easier
- Need to automate scanning and analysis
- Need more quantitative 3D information

Three key features



High-throughput

P4

- Rapid scanning speed
- Parallel processing for scanning and analysis

Simple scanning and analysis settings

P5

- Easy setting
- Intuitive user interface
- Complete scanning & analysis software in one package

Enhanced realism and accuracy in 3D fluorescence imaging

P6

- Real-time background separation
- Seamless 3D fluorescence images



New technology using light-sheet optics

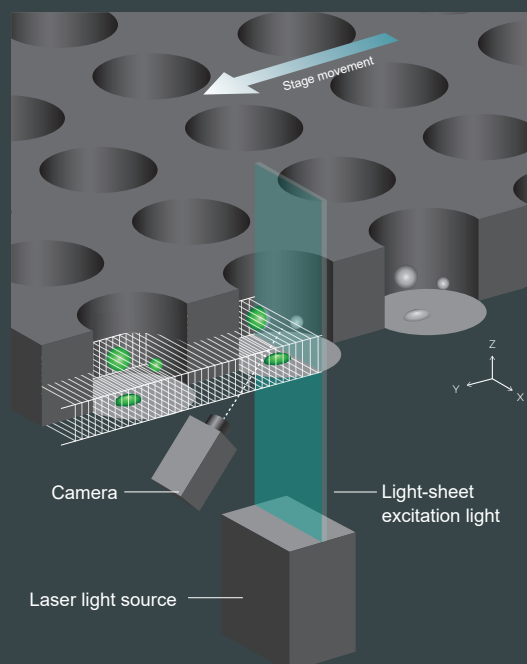
The patented Zyncscan® technology developed by Hamamatsu Photonics is a new technology that combines optical and image processing technologies based on light-sheet optics.

The light-sheet optics of Zyncscan® technology emit a sheet of excitation light from directly below the sample and capture XZ tomographic fluorescence images from an oblique downward direction. By continuously acquiring XZ tomographic fluorescence images while moving the microplate in a direction orthogonal to the light-sheet, 3D fluorescence images are constructed within a few minutes per color.

In parallel with the acquisition of XZ tomography fluorescence images, background fluorescence is removed in real time by three-dimensionally separating background fluorescence and sample fluorescence from the actual XZ tomographic fluorescence image acquired.

Advantages of Zyncscan®

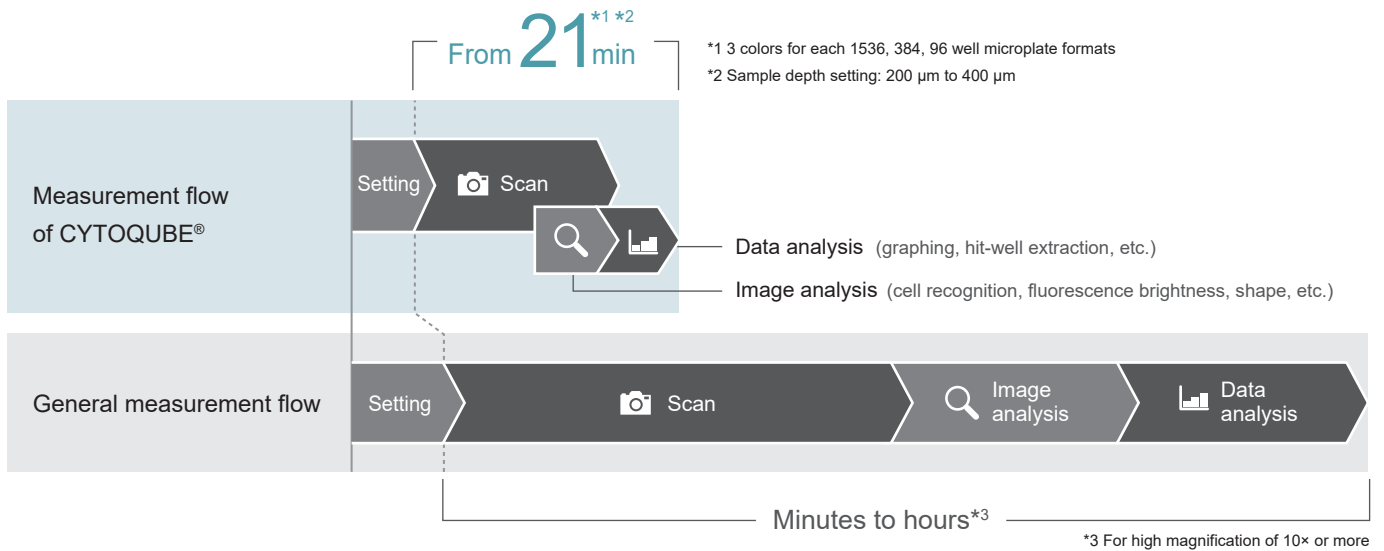
- High-speed scanning of whole wells in a wide field of view
- Z-axis optical performance equivalent to 10× confocal
- Focus free
- Real-time background separation
- Seamless 3D fluorescence images





High-throughput

Measurement flow



Rapid scanning speed

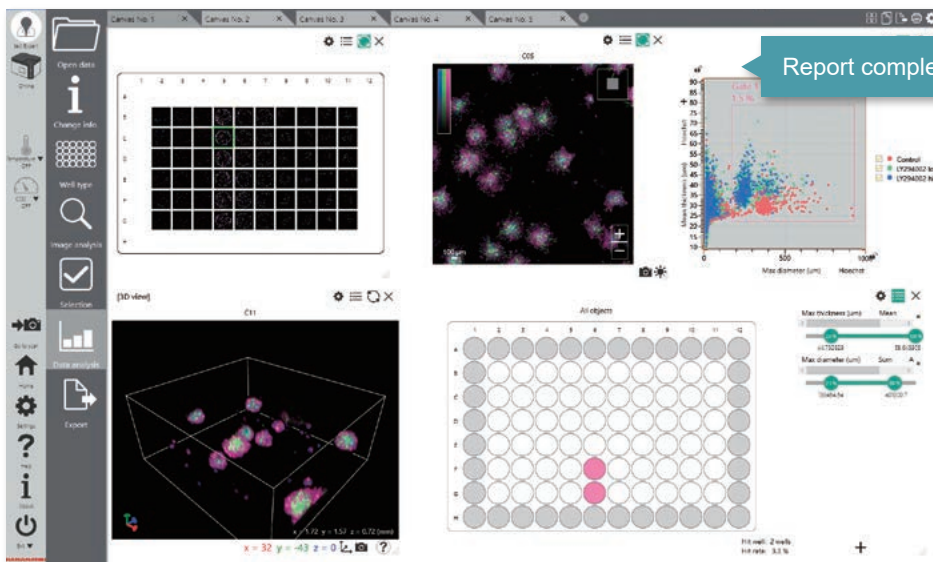
Acquire whole-well and whole-plate fluorescence images within a few minutes per color, regardless of plate format (1536, 384, 96 wells) or sample depth.

	Scanning to analysis time ^{*1}	
	2D sample ^{*2}	3D sample ^{*3}
1 color	4 min to 11 min	7 min to 11 min
2 colors	8 min to 22 min	14 min to 22 min
3 colors	12 min to 34 min	21 min to 34 min
4 colors	16 min to 45 min	28 min to 45 min

Parallel processing for scanning and analysis

The parallel processing of fluorescence image construction and image analysis has significantly shortened the time from scan start to data analysis result output.

*1 Whole plate measurement
*2 Sample depth setting: 0 µm
*3 Sample depth setting: 400 µm



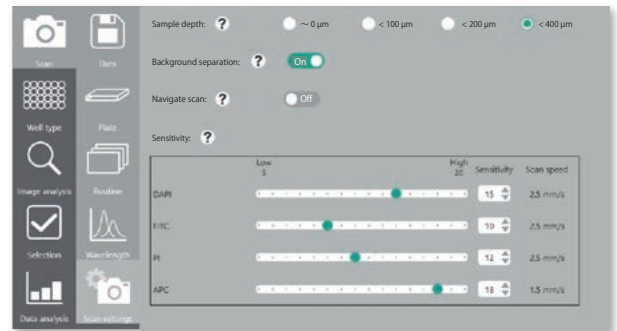
Simple scanning and analysis settings

Easy setting

The settings have been simplified to make it easy to use. Unnecessary settings such as 'focus adjustment' or 'exposure time adjustment' are not included. The optimal sensitivity can be set automatically with the Navigation Scan function. Additionally, you can save the entire process from scanning to result display as a protocol file, making it easy to reproduce scanning and analysis conditions.

Only three settings are required for scanning:

1. Sample depth (4 choices)
2. Background separation (ON or OFF)
3. Sensitivity (can be set automatically)



Intuitive user interface

Settings required for image analysis can be set intuitively, as necessary settings are summarized for each application.

Example image analysis settings:



Find objects/Spheroid

Fluorescence intensity and 3D morphological information (height, thickness, volume) measurements of individual cells and individual spheroids/organoids are analyzed and results are displayed.



Cell invasion

The well is divided into multiple regions, and measurements are performed in each region.



Neighboring cells

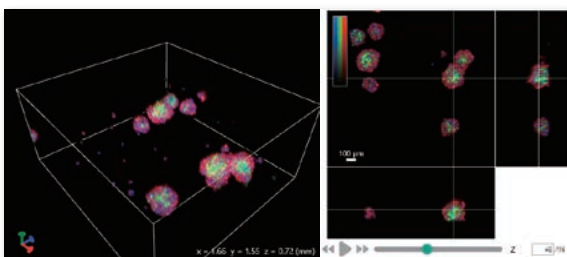
Find the number of adjacent cells and spheroids/organoids.



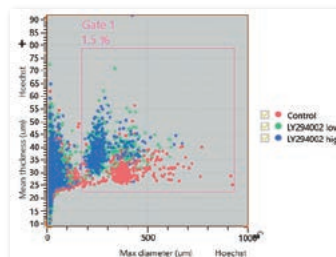
Complete scanning & analysis software in one package

A single software can be used to check 2D/3D fluorescence images of each well, calculate EC50/IC50, create various graphs, identify hit wells, and generate simple reports.

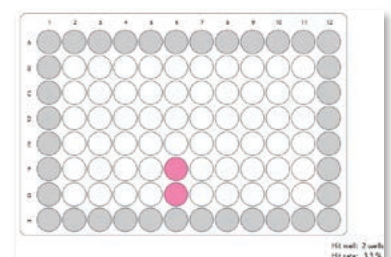
3D fluorescent image display



Scatter plots & gate settings



Hit-well identification



Enhanced realism and accuracy in 3D fluorescent imaging

Real-time background separation

Background fluorescence, derived from autofluorescence of the culture medium or fluorescent dyes in the solution, and fluorescence from the sample are separated from the acquired XZ tomographic fluorescence images, in real-time, effectively removing only the background fluorescence in three dimensions.

No medium wash-out needed

A549 cells cultured with DMEM medium, containing serum and fluorescent dye, were measured both in a fluorescence microscope and with CYTOQUBE® to compare the images. Fluorescent dyes: Annexin V-Alexa Fluor 488

Fluorescence microscope

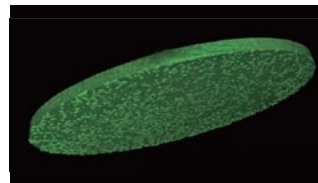


CYTOQUBE®
(background separation ON)

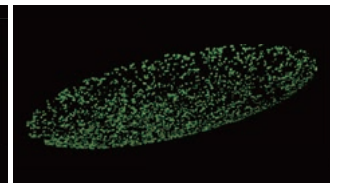


Separation of background signal at ultra-high level

Real-time background separation
OFF



Real-time background separation
ON

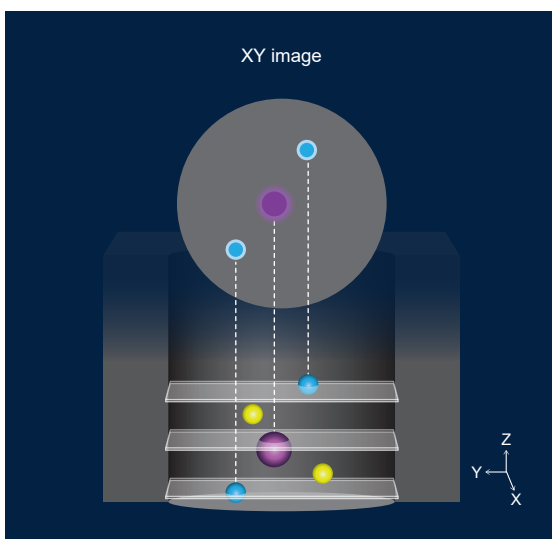


Seamless 3D fluorescent images

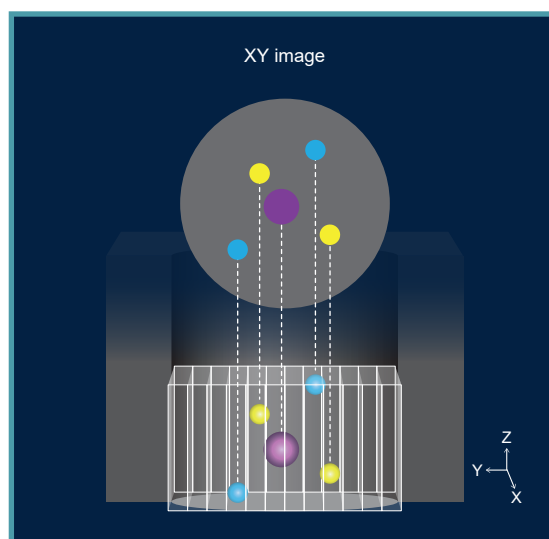
In a typical confocal microscope, multiple 2D XY images at different depths are acquired to construct a 3D fluorescence image. To obtain more accurate 3D information, a larger number of image acquisitions is required.

CYTOQUBE® consistently provides seamless 3D fluorescence images using Zyncscan® technology, ensuring precise 3D information without the need for extensive image acquisition.

Typical confocal microscope



CYTOQUBE®



Applications

3D samples

Spheroids, organoids, patient-derived tissue samples, bioprinting samples, etc.

2D samples

Established cells, primary cultured cells, differentiated cells derived from iPS cells, etc.



Migration/Invasion assay



Morphological analysis



Analysis of the number of adjacent cells



Live/dead assay



Reactive Oxygen Species (ROS) measurement



Apoptosis assay



Angiogenesis assay

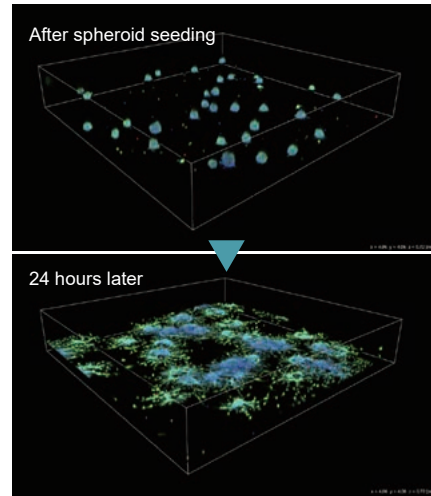


Fluorescence measurement of cell surface markers



Migration/Invasion assay

Cancer cell (U-87 MG) spheroids of approximately 200 μm diameter were seeded into a 96 well microplate and cultured in a medium containing Matrigel. After 24 hours, we observed the migration/infiltration of cells from the spheroids.

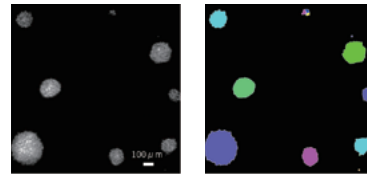


Hoechst 33342 (blue), Calcein (green), Propidium Iodide (red)



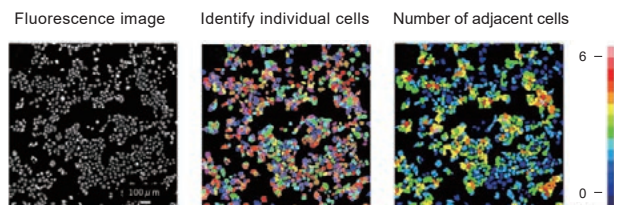
Morphological analysis

The brightness, diameter, volume etc. of individual cells, spheroids and organoids can be quantified.



Analysis of the number of adjacent cells

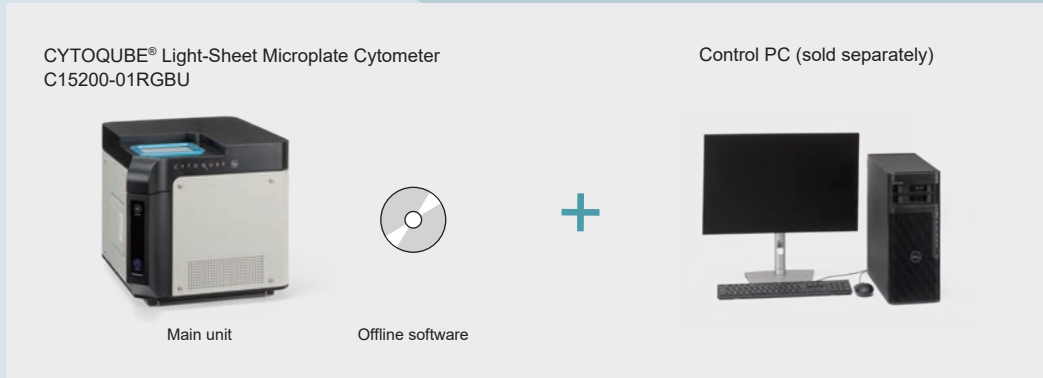
The analysis assesses the quantity of neighboring cells, including their fluorescence intensity, brightness, area, etc. These measured values enable the evaluation of cell-to-cell contact and aggregation.



Scan here for application notes and other information.



System configuration



Specifications

CYTOQUBE® Light-Sheet Microplate Cytometer

Type number	C15200-01RGBU	
Scanning method	High-speed 3D fluorescence measurement method using light-sheet illumination (Zyncscan®)	
Excitation laser	Select from 405 nm, 488 nm, 561 nm and 637 nm (up to 4 types can be selected)	
Fluorescence filter	Select from 458 nm, 531 nm, 600 nm and 680 nm (up to 6 sets can be installed)	
Imaging device	Scientific CMOS image sensor	
Image resolution	2.75 μm (X) × 2.75 μm (Y) × 6.215 μm (Z)	
Maximum observable height ^{*1}	400 μm	
Focus	Focus free (Zyncscan®)	
Microplate	1536, 384, 96 well microplates (SBS standard format)	
Setting environment	Temperature setting range	+30 °C to +45 °C (ambient temperature: +25 °C)
	CO ₂ concentration monitoring range	0 % to 10 % (with CO ₂ inlet)
Barcode reader	1D barcode (reading position: both sides of the short side of the microplate) ^{*2}	
Power supply	AC 100 V to AC 240 V	
Power consumption	Approx. 700 VA	
Operating environment	Ambient operating temperature	+20 °C to +30 °C
	Ambient operating humidity	30 % to 80 % (with no condensation)

^{*1} Height from the bottom of the well when the flatness of the bottom of the microplate is ± 50 μm in the short side direction

^{*2} 1536, 384 and 96 plates less than 9.5 mm thick may not read properly.

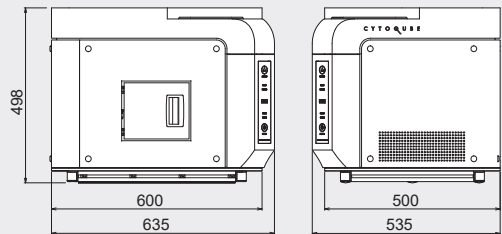
^{*} For automation function, please contact our sales representatives.

CYTOQUBE® software

Type number	U15134-01	
Analysis item (at each fluorescence wavelength of individual cells / spheroids)	Fluorescence intensity per unit volume Morphology of cellular objects (diameter, thickness / height, volume)	
Display item	Data	Table, heat well map, hit well map, 2D dot plot, histogram, dose response curve
	Image	2D image (at well unit or whole plate), 3D image (at well unit)
Data output format	Measurement data (.csv), snapshot image (.tiff / .jpeg / .bmp / .png), 3D movie (.avi / .wmv), original image (.tiff)	

Dimensional outline (Unit: mm)

Weight: Approx. 70 kg
(Main unit when 4 types of excitation lasers are installed)



Laser safety

CYTOQUBE® is classified as a Class 1 laser product, conforming to international laser safety standards (USA: 21 CFR 1040.10 CDRH, other areas: IEC 60825-1). These standards mandate manufacturers to provide preventive safety measures. Hamamatsu lasers are appropriately classified, and they come with necessary safety measures and labeling. During operation, users must also adhere to preventive safety measures as per laser-related regulations.



Description Label (Sample)



Caution Label

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 - The product described in this brochure is designed to meet the written specifications, when used strictly in accordance with all instructions.
 - Specifications and external appearance are subject to change without notice.
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